

Measurement and Partitioning of *in situ* Carbon Fluxes in Turfgrasses

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Introduction:

Field measurements of photosynthesis and respiration are often conducted with surface chambers that cover a small area of the turf's canopy. Consequently, measurements include the various components of ecosystem carbon (C) balance, including photosynthesis, canopy respiration and soil respiration. When used for measuring carbon flux, steady-state portable photosynthesis systems (i.e., Licor 6400) are pressurized. Small changes in pressurization are known to partially suppress soil respiration. Thus, chamber measurements of CO₂ flux from turf may be more related to unintended changes in soil respiration than to changes in photosynthesis. We propose that the chamber can be intentionally pressurized to almost completely suppress soil respiration and obtain independent measurements of gross canopy photosynthesis and canopy respiration.

Objectives:

1. Fabricate and test a chamber connected to a portable photosynthesis system where chamber pressure could be manipulated (Fig 1).
2. Apply increasing pressure until soil respiration was prevented from entering the chamber. The result would be a more accurate indicator of treatment effects on photosynthesis and canopy respiration.
3. Attempt to partition various components in the turf C budget, including net and gross photosynthesis and canopy respiration, in perennial ryegrass, tall fescue, and Kentucky bluegrass.

Theory:

The instantaneous C budget (CO₂ fluxes) in turfgrass can be represented by:

$$NEE = P_g - R_c - R_s$$

where *NEE* is net ecosystem exchange of CO₂, *P_g* is gross photosynthesis, *R_c* is canopy respiration, and *R_s* is soil respiration.

If *R_s* could be completely suppressed with pressure, then sunlit and shaded measurements on the same area would yield (*P_g*-*R_c*) and *R_c*, respectively. The difference between the two readings would be *P_g*. However, field testing showed that although *R_s* could be radically reduced with intentional over pressurization, it could not be completely eliminated. Thus, an alternative procedure was developed that accounted for this small soil flux.

Materials and Methods

- Licor 6400 with modified custom chamber (Fig. 1).
- Measurements collected at Rocky Ford Turfgrass Research Center, Manhattan, KS.
- Soil type: silt loam
- Gravimetric soil water content collected from 0-10 cm.
- Chamber measurements of CO₂ flux were collected from intact turf canopies under full sunlight, shaded (eliminated photosynthesis), and after turf was clipped at ground level (eliminated *R_c*).

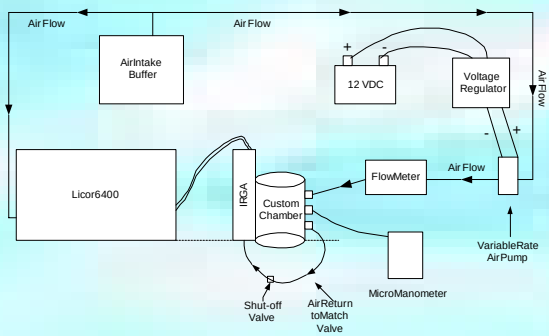


Figure 1. Conceptual diagram (top) and photo (bottom) that illustrate the design of a pressurized chamber system. A variable flow air pump and manometer were used to regulate chamber pressure. A mass flow meter was used to monitor the air flow into the chamber, and results were used to correct fluxes for the effect of dilution.

Results:

Preliminary measurements indicated that:

1. Neutral pressure chamber measurements were extremely difficult to obtain in an open-path system because changes in pressure of ±0.5 Pa were often uncontrollable and caused large differences in *R_s* (Fig. 2); hence, measurements of *NEE* were not measured in this study.
2. Chamber pressurization prevented most but not all *R_s* from entering the chamber (Fig. 3). This problem was circumvented by measuring CO₂ flux from a clipped, shaded area near the plots under investigation. This provided an estimate of *R_s'*, the soil flux when the chamber was under pressure.

The following three measurements from fully pressurized chambers provided estimates of the given components of the C budget:

1. Sunlit chamber: *P_g*-*R_c*-*R_s'*
2. Shaded chamber: *R_c*-*R_s'*
3. Turf canopy clipped at ground level: *R_s'*

Therefore, estimates of net photosynthesis (*P_{net}*), *P_g*, and *R_c* could be calculated from the above measurements:

1. *P_g* = Sunlit chamber – shaded chamber
2. *P_{net}* = Sunlit chamber – clipped canopy*
3. *R_c* = Shaded chamber – clipped canopy*

*Measurements collected immediately after turfgrass canopy was clipped.

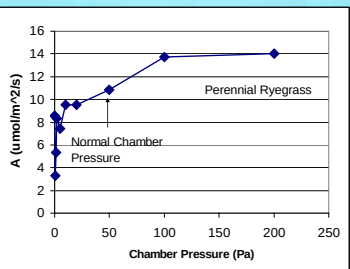


Figure 2. Apparent response of carbon assimilation (*A*) to changes in chamber pressure (sunlit). The increase in CO₂ “assimilation” with chamber pressure was actually caused by reductions in *R_s* entering the chamber.

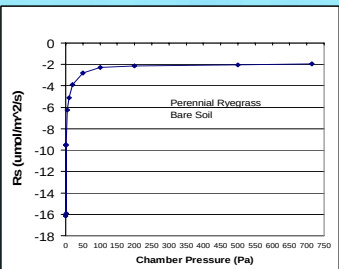


Figure 3. Response of soil respiration (*R_s*) entering the chamber to changes in chamber pressure (bare soil). Of the total *R_s* of 16 umol m⁻² s⁻¹, all but 1.92 umol m⁻² s⁻¹ was excluded by high pressures.

Results (continued):

- Measurements of CO₂ fluxes increased by 6 to 23% when the chamber was pressurized compared to normal chamber operation (Fig. 4).
✓ Increase was result of higher pressure restricting *R_s* from chamber measurements.
- Estimates of *P_g*, *P_{net}*, and *R_c* provided more meaningful quantifications of C fluxes because these components are directly linked to the biophysics of the turf canopy, as opposed to normal operation of the chamber, where the partitioning of C fluxes among *P_g*, *P_{net}*, and *R_c*, is uncertain (Fig. 5).
- Measurements of C fluxes at the beginning and end of a one-week drydown showed a 19 and 11% reduction in *P_g*, a 21 and 16% reduction in *P_{net}*, and an 11% reduction and 10% increase in *R_c* in perennial ryegrass and tall fescue, respectively (Fig. 5).
✓ Comparatively, measurements with the unmodified chamber showed a 26 and 21% reduction in C fluxes in perennial ryegrass and tall fescue, respectively.

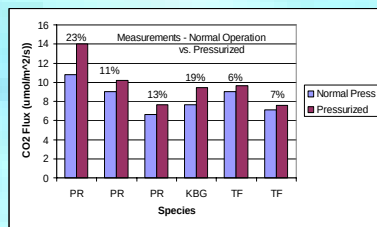


Figure 4. Comparison between measured C fluxes with normal chamber operation (unmodified) and with pressurized chamber (modified) in perennial ryegrass (PR), Kentucky bluegrass (KBG), and tall fescue (TF). PR and TF were measured on multiple dates.

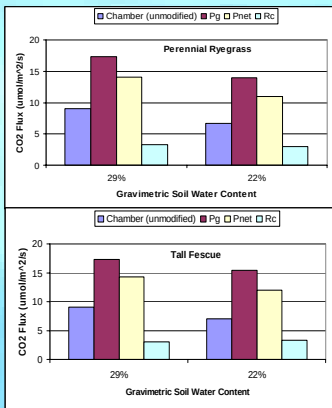


Figure 5. Carbon fluxes measured with normal (unmodified) and modified chamber on perennial ryegrass and tall fescue during a one-week drydown. Variables *P_g*, *P_{net}*, and *R_c* were partitioned by using modified chamber on sunlit, shaded, and clipped turfgrass.

Conclusions:

1. Measuring CO₂ fluxes from turf using an open-chamber system that does not account for pressure yields meaningless results that do not represent *NEE* or *P_g*-*R_c*.
2. C fluxes can be partitioned among *P_g*, *P_{net}*, and *R_c* using a modified, pressurized chamber consecutively on sunlit, shaded, and clipped turfgrass.
3. Partitioning of C fluxes provided more sensitive and meaningful comparisons between treatments than measurements with the unmodified chamber because of the direct relationship of *P_g*, *P_{net}*, and *R_c* to the biophysics of the turfgrass canopy.



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