

The effect of altered rainfall patterns on leaf rust severity in tallgrass prairie

J. K. McCarron (1), K. A. Garrett (1), S. P. Dendy (1), Z. Su (1), P. A. Fay (1), H. M. Alexander (2) and B. M. Broeckelman (1) Kansas State University, Manhattan; (2) University of Kansas, Lawrence.

Abstract

The altered precipitation patterns predicted by climate change models will likely affect the incidence and severity of plant diseases in natural systems. We studied the effects of precipitation patterns on the pathogens of 23 common tallgrass prairie plant species at Konza Prairie Biological Station, finding a decrease in infection with decreasing precipitation for most, but not all, species. We studied in more detail the effects of precipitation patterns on leaf rust caused by *Puccinia dioicae* on goldenrod, *Solidago canadensis*, a common native forb. Precipitation is important for typical rust pathogens in two distinct seasons of the year. In spring, adequate leaf wetness duration is necessary for infection of the alternate host during a limited window of opportunity. In summer, greater numbers of intervals of adequate leaf wetness duration will allow for more generations of pathogen increase on the primary host, which allow for more spores to over winter.

Introduction

Changes in precipitation patterns may have important effects on plant pathogens in tallgrass prairie. Many foliar pathogens depend on leaf surface wetness for successful infection, but model predictions include reduced rainfall and greater intervals between rain events (Easterling 1990, Houghton et al. 1990, 1996, Karl et al. 1991). Two experiments at Konza Prairie Biological Station (KPBS) near Manhattan, Kansas, have been designed to study the long-term effects of altered patterns of precipitation on a mesic tallgrass prairie ecosystem (Fay et al. 2000). Plants may experience a trade-off as water becomes more limiting: they may experience water stress but they may also experience less infection by pathogens.

In our studies at KPBS so far, rust fungi have proven to be the most common foliar pathogens that produce clear visual symptoms. Many rust fungi have complex life cycles that include an alternate host where sexual reproduction takes place and a primary host where asexual reproduction takes place. The alternate host is infected in spring and the asexually produced on the alternate host can only infect the primary host. Urediniospores produced on the primary host can re-infect the primary host species to produce multiple generations of infection over the course of the summer. As primary hosts begin to senesce, or if primary hosts experience stress, the rust fungi switch from producing urediniospores to teliospores, which cannot re-infect the primary host but are adapted to overwinter. (Arthur 1962). Because of this complex life cycle, rust fungi require adequate moisture at two different important points in the year. In spring, adequate moisture is required for the one chance at sexual reproduction. In summer, the greater the period during which adequate moisture is available, the more generations of reproduction are possible. If plants experience extreme drought stress at any point in the summer, the fungi may switch to producing teliospores so that no more fungal generations are possible that year.

Acknowledgments
 • John W. McClain for identification of *Puccinia dioicae*
 • Tom Sweeney for technical support
 • This work was supported by the National Science Foundation under Grant No. DEB-01-10033 and by NSF Grant No. EPS-04-54732 with matching support from the State of Kansas

Response of foliar disease severity of common tallgrass prairie species to water stress

An irrigation experiment at KPBS initiated in 1990 addresses the effects of steady high levels of moisture on prairie plants (Knapp et al. 2001). Paired irrigation and control transects span upland and lowland topographic positions in annually burned prairie. Irrigation is conducted following a protocol designed to offset potential water deficits based on the Penman-Monteith model estimates of evapotranspiration, natural precipitation inputs, and soil water potentials. We estimated foliar disease severity in the lowland transects of this experiment for a set of 23 common tallgrass prairie plant species (Table 1) in 2001, and ongoing sampling is in progress.

Some plant species had no foliar infection with clear visual symptoms. Many species were infected by rust fungi (Table 1). Disease severity for the majority of these host-pathogen combinations was lower in the absence of irrigation, though there were some exceptions (Fig. 1). Our ongoing work in this experiment will allow us to determine whether the exceptional host-pathogen combinations show a consistent difference in response to changes in precipitation. Different responses to changes in precipitation by different host-pathogen combinations may contribute to greater stability in productivity in response to environmental change for more diverse plant communities.

Species	Common name	Pathogen	Primary alternate host (infection in spring)
1	Blue grama	None	None
2	Dark green needlegrass	None	None
3	Little bluestem	None	None
4	Red top	None	None
5	Yellow bluestem	None	None
6	Blue grama	None	None
7	Red top	None	None
8	Yellow bluestem	None	None
9	Blue grama	None	None
10	Red top	None	None
11	Yellow bluestem	None	None
12	Blue grama	None	None
13	Red top	None	None
14	Yellow bluestem	None	None
15	Blue grama	None	None
16	Red top	None	None
17	Yellow bluestem	None	None
18	Blue grama	None	None
19	Red top	None	None
20	Yellow bluestem	None	None
21	Blue grama	None	None
22	Red top	None	None
23	Yellow bluestem	None	None
24	Blue grama	None	None
25	Red top	None	None
26	Yellow bluestem	None	None
27	Blue grama	None	None
28	Red top	None	None
29	Yellow bluestem	None	None
30	Blue grama	None	None
31	Red top	None	None
32	Yellow bluestem	None	None
33	Blue grama	None	None
34	Red top	None	None
35	Yellow bluestem	None	None
36	Blue grama	None	None
37	Red top	None	None
38	Yellow bluestem	None	None
39	Blue grama	None	None
40	Red top	None	None
41	Yellow bluestem	None	None
42	Blue grama	None	None
43	Red top	None	None
44	Yellow bluestem	None	None
45	Blue grama	None	None
46	Red top	None	None
47	Yellow bluestem	None	None
48	Blue grama	None	None
49	Red top	None	None
50	Yellow bluestem	None	None
51	Blue grama	None	None
52	Red top	None	None
53	Yellow bluestem	None	None
54	Blue grama	None	None
55	Red top	None	None
56	Yellow bluestem	None	None
57	Blue grama	None	None
58	Red top	None	None
59	Yellow bluestem	None	None
60	Blue grama	None	None
61	Red top	None	None
62	Yellow bluestem	None	None
63	Blue grama	None	None
64	Red top	None	None
65	Yellow bluestem	None	None
66	Blue grama	None	None
67	Red top	None	None
68	Yellow bluestem	None	None
69	Blue grama	None	None
70	Red top	None	None
71	Yellow bluestem	None	None
72	Blue grama	None	None
73	Red top	None	None
74	Yellow bluestem	None	None
75	Blue grama	None	None
76	Red top	None	None
77	Yellow bluestem	None	None
78	Blue grama	None	None
79	Red top	None	None
80	Yellow bluestem	None	None
81	Blue grama	None	None
82	Red top	None	None
83	Yellow bluestem	None	None
84	Blue grama	None	None
85	Red top	None	None
86	Yellow bluestem	None	None
87	Blue grama	None	None
88	Red top	None	None
89	Yellow bluestem	None	None
90	Blue grama	None	None
91	Red top	None	None
92	Yellow bluestem	None	None
93	Blue grama	None	None
94	Red top	None	None
95	Yellow bluestem	None	None
96	Blue grama	None	None
97	Red top	None	None
98	Yellow bluestem	None	None
99	Blue grama	None	None
100	Red top	None	None

Table 1. Plant species studied in disease transects illustrated in Fig. 1, along with other potential primary and alternate hosts for the pathogen.

Treatment	Amount of Precipitation	Interval between Applications	Years
Control	Natural Amount	Natural Interval	1999-2003
Drought	Decreased by 30%	Natural Interval	1999-2003
Increased Interval	Natural Amount	Increased by 50%	1999-2003
Drought + increased interval	Decreased by 30%	Increased by 50%	1999-2003

Table 2. Rainfall manipulation treatments in the RAMP project



Fig. 1. Photos of *Solidago* rust and RAMP project

Effects of precipitation change on incidence of infection by the rust fungus *Puccinia dioicae*

We have studied *P. dioicae* for four years in the context of an experiment at KPBS designed to address the effects of decreased precipitation and increased intervals between precipitation events (Knapp et al. 2001). We estimated disease incidence in *Solidago canadensis* (Canada goldenrod), an alternate host of this pathogen that is infected in May. The primary hosts of *P. dioicae* include a number of *Carex* sp that are infected during the summer. Rainfall exclusion shelters were erected over 12 6x6 m plots (Fig. 4). The shelters were constructed with a steel tube frame and a transparent plastic roof. Each shelter had a rainfall collecting system, storage tanks and a sprinkler system for re-applying rainfall. Rainfall was collected and re-applied according to the protocol in Table 2. There were three replicates in a randomized complete block design. We sampled three quadrats totaling 4 m² in each plot.

The treatments imposed in this experiment had varying effects during 2000-2004 (Fig. 5). In 2000, the pattern was as we might have expected and the differences were statistically significant ($p < 0.05$). In 2001 incidence was very low and no differences due to the treatments were observed; instead, there was some evidence in this year for an effect of primary host abundance within plots on incidence on *S. canadensis*. In 2002 disease incidence was very high and no treatment effects were observed; in this year replacement of the shelters in spring was delayed so some infection may have occurred before the shelters were in place. In 2003 incidence was again low and no clear treatment effect was observed.

We also considered the results in the untreated plots as a function of the ambient weather conditions over the four years (Fig. 6). We interpreted the precipitation pattern using the estimated LWD described above (Fig. 3). Based on the life cycle of *P. dioicae*, we would expect that two time segments are important to pathogen abundance. In May, the availability of more potential infection intervals should increase the success rate for infection of *Solidago* per propagule produced in the previous summer. The number of propagules available in May should be determined by the number and timing of potential infection intervals in the previous summer when *Carex* was infected. The years 2000, 2002, and 2003 had similar patterns of spring moisture availability and the differences in incidence were consistent with the differences in the number of potential weeks of pathogen population increase during the previous summer. In 2001 incidence was lowest of all; this year had the greatest number of potential weeks of pathogen population increase during the previous summer, but had the driest spring.

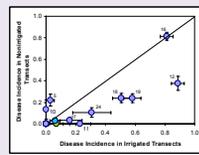


Figure 1. Pathogen load response to change in precipitation. Each point indicates the proportion leaves infected for a different one-plant species (see Table 1 for species number key). Standard errors are indicated (green oval in quadrants number 1, blue oval in quadrants number 2).

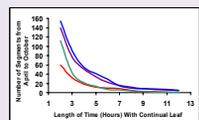


Figure 2. The number of potential infection intervals during the growing season for the Konza Prairie Biological Station in northeast Kansas (39°51'N, 96°37'W). Leaf wetness duration was estimated for pathogens requiring at least 4-hour intervals. (May = week 15-21 and summer = week 22-23)

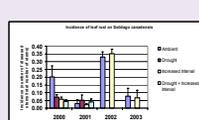


Figure 3. The estimated number of free intervals of sufficient leaf wetness occurring during the growing season (LWD) as a function of the minimum leaf wetness duration (LWD) required for infection. These four years illustrate the importance of conditions of years, especially for pathogens with other LWD requirements.

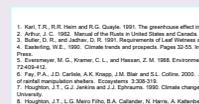


Figure 4. Disease incidence on *Solidago canadensis* in RAMP plots from 2000 to 2003. Bars represent the incidence on disease number of observed quadrats (number of stems x standard error). Note that only two treatments were applied in 2000-2003.

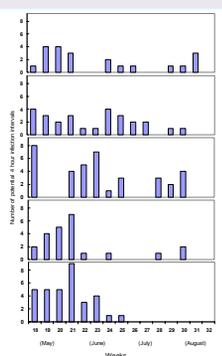


Figure 5. Disease incidence on *Solidago canadensis* (indicated by bubble size) as a function of the potential number of infection weeks in previous summer and current May.

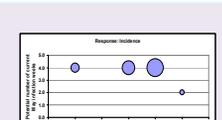


Figure 6. Disease incidence on *Solidago canadensis* (indicated by bubble size) as a function of the potential number of infection weeks in previous summer and current May.

1. Karl, T.R., R.H. Heins and R.G. Quayle. 1991. The greenhouse effect in central South America: Is it real, when? Science 251:1058-1061.
 2. Arthur, F. C. 1962. *Journal of the Royal Microscopical Society and Canada*. Oxford: Blackwell.
 3. Easterling, W.E. 1990. Climate trends and prospects. Pages 33-55. In R.N. Sarason and D. Hare, eds. *Natural Resources for the 21st Century*. Washington: Island Press.
 4. Eisenmann, M.G., Korman, C.L., and Heenan, J. M. 1988. Environmental Influences on the Establishment of *Puccinia-Rhizoctonia* Infection Structures. Plant Disease 72:420-423.
 5. Fay, P. A., J.C. Garbutt, A.K. Probst, J.B. Blair and S.L. Collins. 2000. Altered rainfall timing and quantity in a mesic grassland ecosystem: Design and performance of rainfall-manipulation shelters. *Ecosystems* 3:306-318.
 6. Houghton, J.T., C.J. Imhoff and J. Squires. 1990. Climate change: the IPCC Scientific Assessment. World Meteorological Association, Cambridge: Cambridge University Press.
 7. Houghton, J.T., L.G. Meira Filho, B.A. Callander, N. Harris, A. Kubiak and M.K. Maskell. 1998. *Climate Change 1998: The Science of Climate Change*. Cambridge: Cambridge University Press.
 8. Hare, L. and J. J. Garbutt. 1992. Modeling Leaf Wetness in Relation to Plant Disease Epidemiology. *Ann. Rev. Phytopathol.* 30:555-577.
 9. Karl, T.R. 2002. Science to enhance the specific identification of leaf wetness duration. *Plant Disease* 86:179-180.
 10. Knapp, A.K., D. Stapp, J. M. and Garbutt, J. K. 2001. Frequency and extent of water limitation to primary production in a mesic temperate grassland. *Ecosystems* 4:19-29.
 11. de Villiers-Pagan, C., Hare, L., Leonard, M. and Collins, H. 1995. Comparative effects of temperature and integrated wet periods on germination, penetration and infection of *Puccinia recondita* on oat leaves. *Phytopathology* 85:538-543.
 12. Viner, P.A., T.A. Brabury and R. Cross. 1980. The biology of Canadian weeds. 43. *Solidago canadensis* L. *Can. J. Plant Sci.* 60: 1393-1403.