

Conservation often focuses on rapidly declining, or critically rare species

- Very expensive
- Often too little, too late...
- May require a "permanent" life-line into the future.



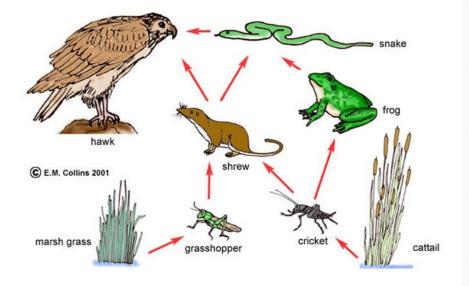
Should we care about imperiled species? What can they tell us?

- They epitomize the problems:
 - Biodiversity declines can be sudden and fast.
 - We normally know very little about rare taxa.
 - Conserving rare taxa is extremely difficult!
- We should care because:
 - They are indicators of change
 - We don't know what role this species fulfilled
 - We need to understand biodiversity connections!



Benefits of studying common species

- High genetic diversity \rightarrow high adaptive capacity
- Large populations \rightarrow easy to work with
- Broad geographic coverage \rightarrow adapted to regional processes
- Lots of intact ecological connections!
 - High potential to resolve them...

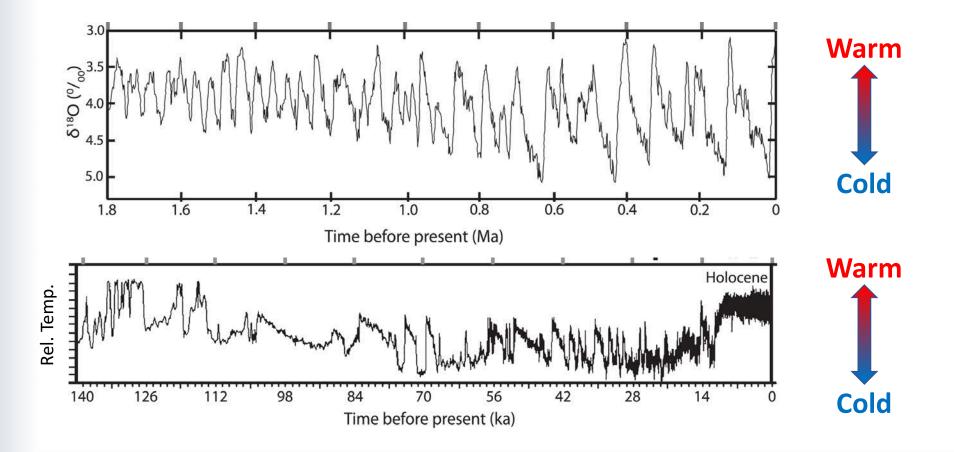


Goals for biodiversity conservation (my view)

- 1. Define the taxonomy Phylogeography
 - Describe species and identify distinct lineages
 - Explore evolutionary processes
- 2. Identify regional centers of diversification
 - Genetic diversity centers/borders over many species?
 - Same spatial/temporal history across species?
 - Regional conservation hotspots?
- 3. Resolve connections among associated biodiversity
 - Community ecology
 - Hosts and parasites Co-evolution.

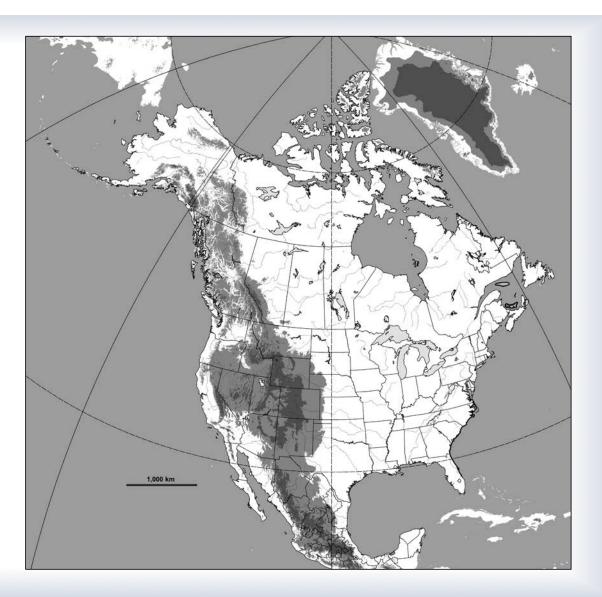
My Study System

Temporal Focus: Quaternary through to present



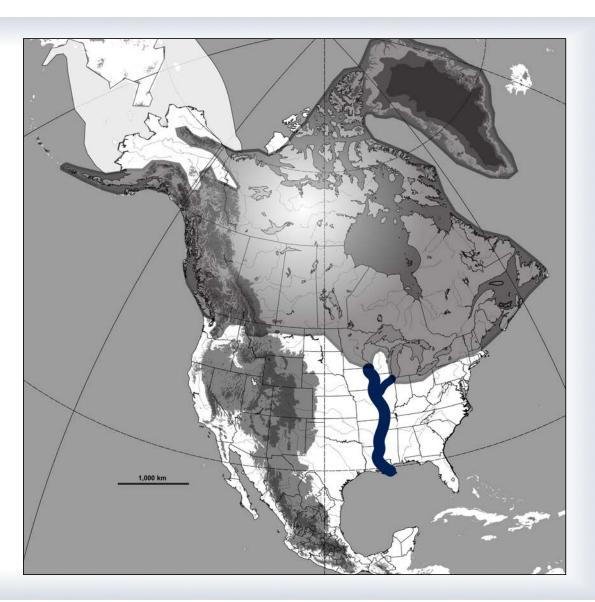
Spatial Focus: Northern hemisphere

Current interglacial



Spatial Focus: Northern hemisphere

Glacial cycle dynamics e.g., America 20 kyr.



Taxonomic focus: Shrews... common, but cool!



The 19 species of shrews found in Canada.

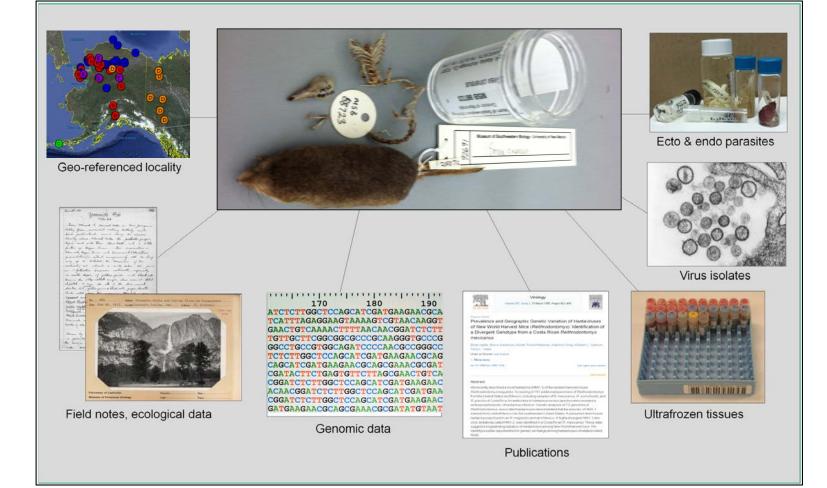
Shrews, genus Sorex ~80 Holarctic species



Resolving generalizable processes of change

- Abundant, ubiquitous
- Tiny, high metabolism, high turnover \rightarrow rapid evolution
- Insectivorous \rightarrow complex parasite life cycles

Evolutionary conservation relies on specimens!!



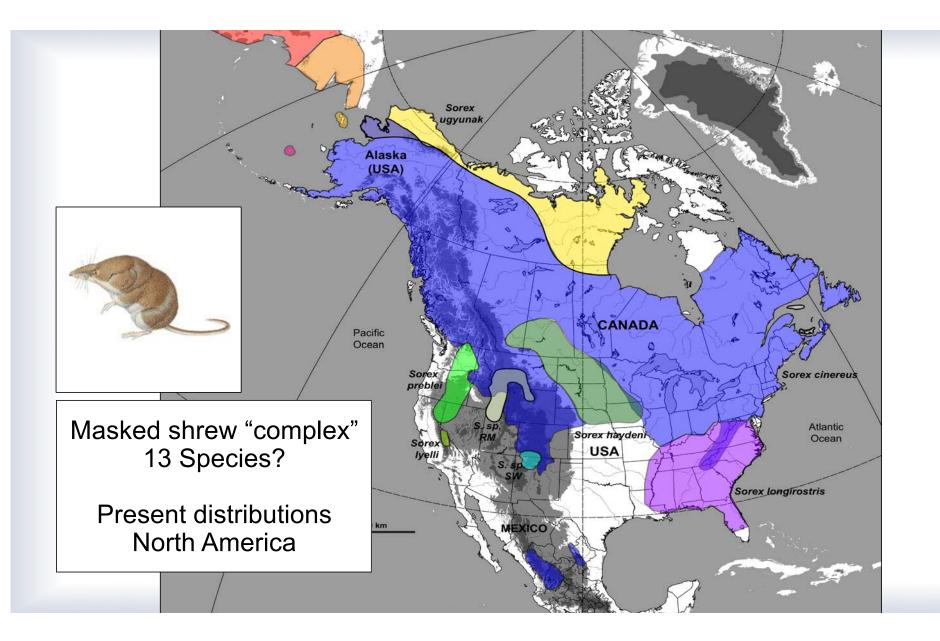
Methods

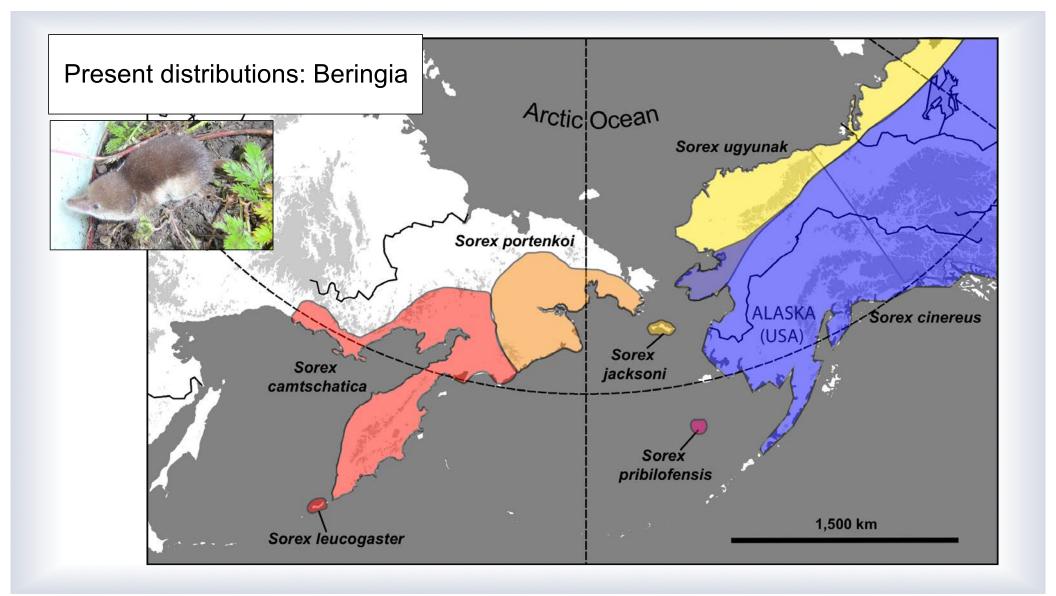
Collect specimens \rightarrow Sequence genes \rightarrow Analyze data

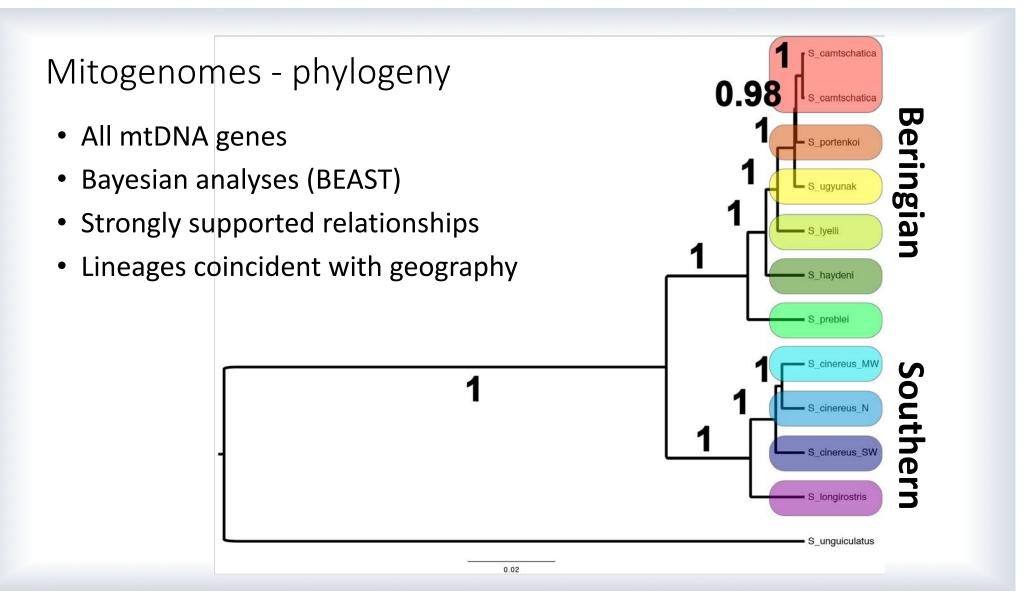
- 800 specimens representing all "species"
- Mitogenome sequencing (in progress)
 - Long-range PCR for ~200 samples
- Nuclear reduced representation sequencing (in development)
 - ~4000 gene regions
- Build phylogenetic trees
- Test for signals of selection adaptation across environments
- Test for hybridization fitness consequences

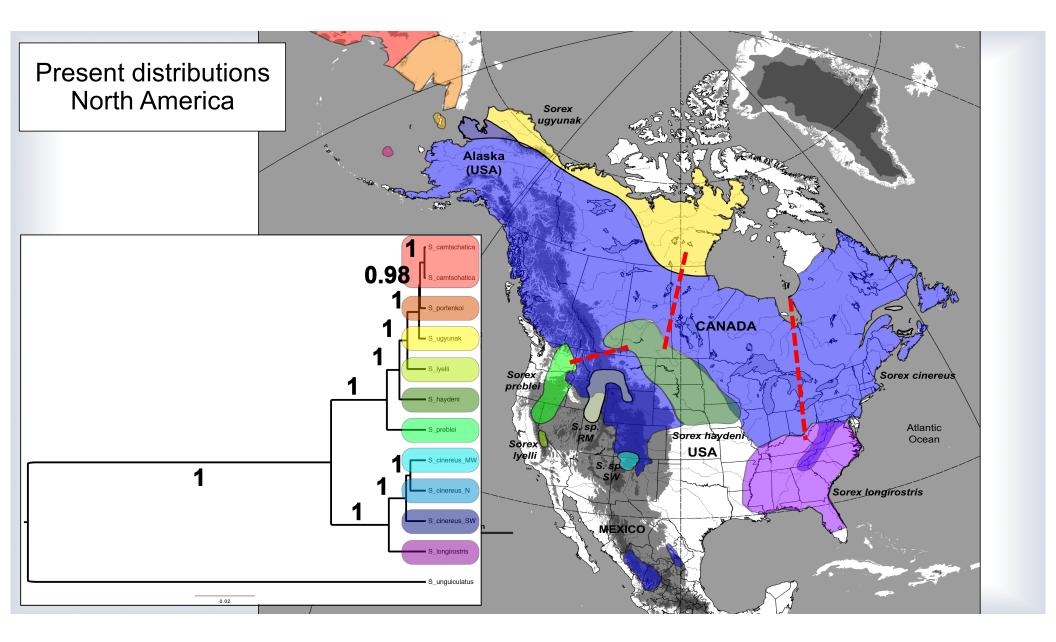
Goals for biodiversity conservation

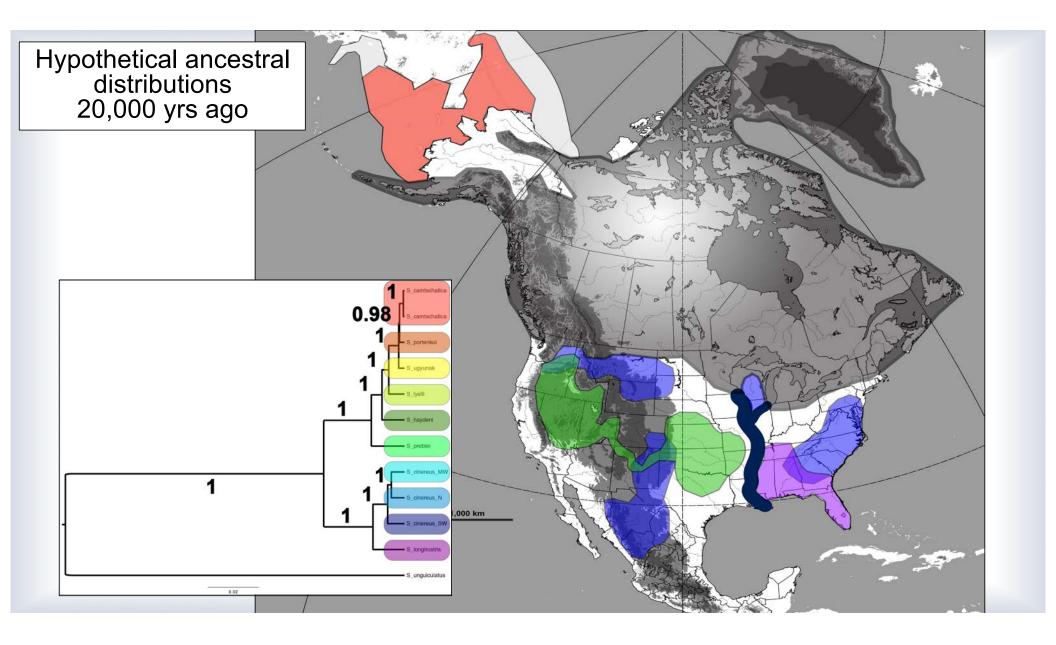
- 1. Define the taxonomy.
 - Describe species and identify distinct lineages
 - Explore evolutionary processes
- 2. Identify regional centers of diversification
 - Genetic diversity centers/borders over many species?
 - Congruent spatial/temporal history across species?
 - Regional conservation hotspots?
- 3. Resolve connections among associated biodiversity
 - Community ecology
 - Hosts and parasites Co-evolution.





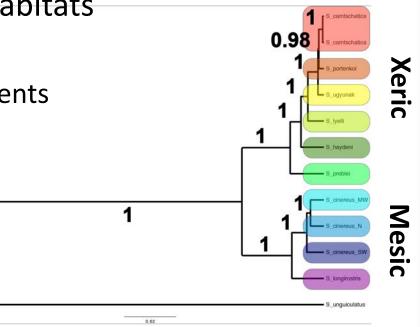


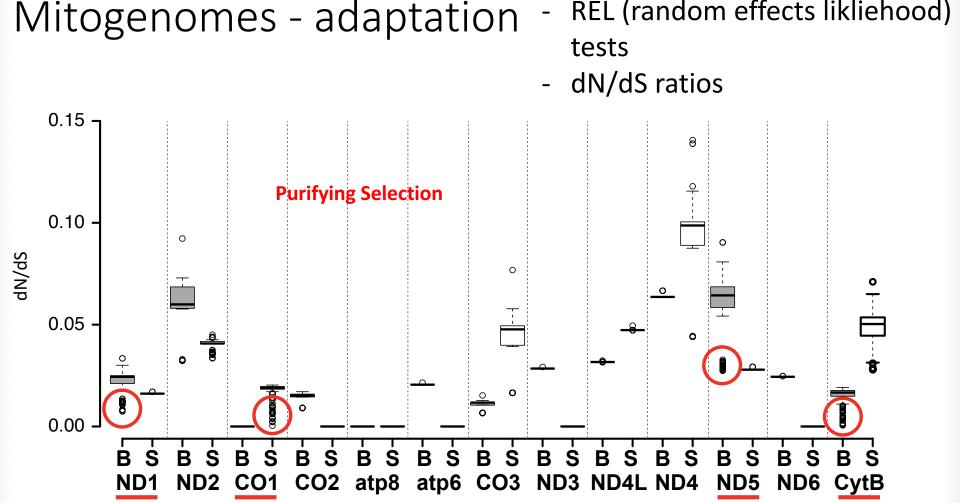




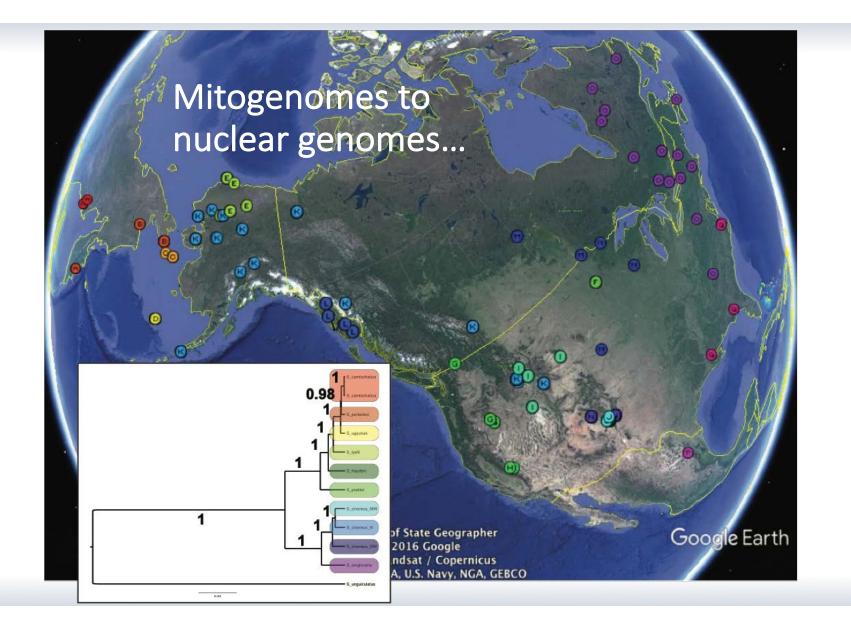
Mitogenomes – adaptation to local environments

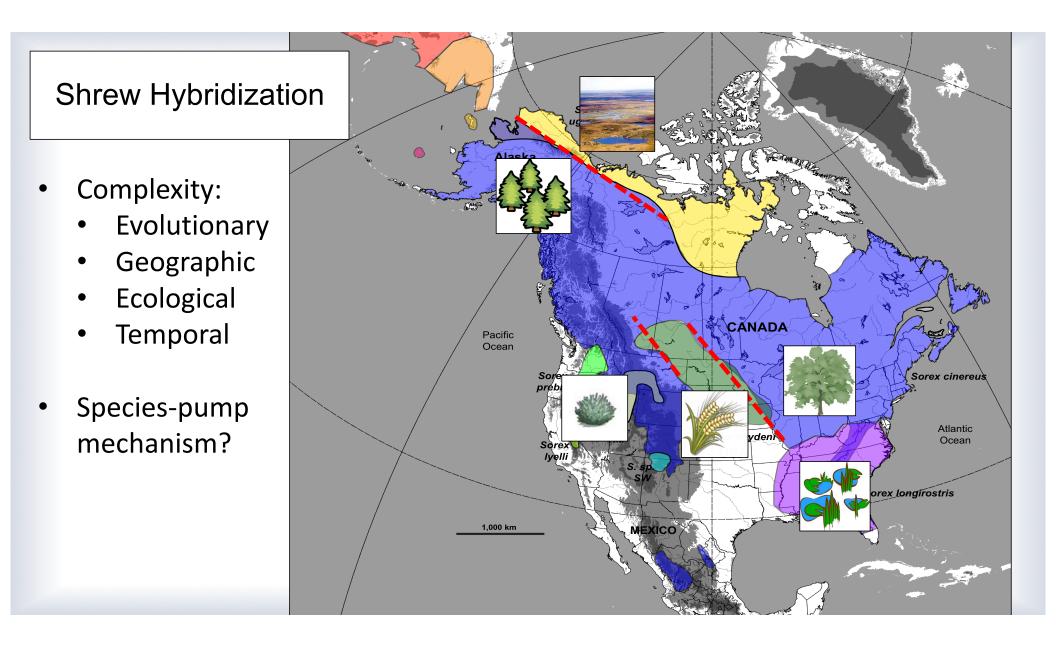
- Masked shrews extremely high metabolism
- Two major clades mesic or xeric habitats
- Expectations:
 - selection across environmental gradients
 - intensified in harsher environments
 - purifying or diversifying selection
 - reflects demographic history

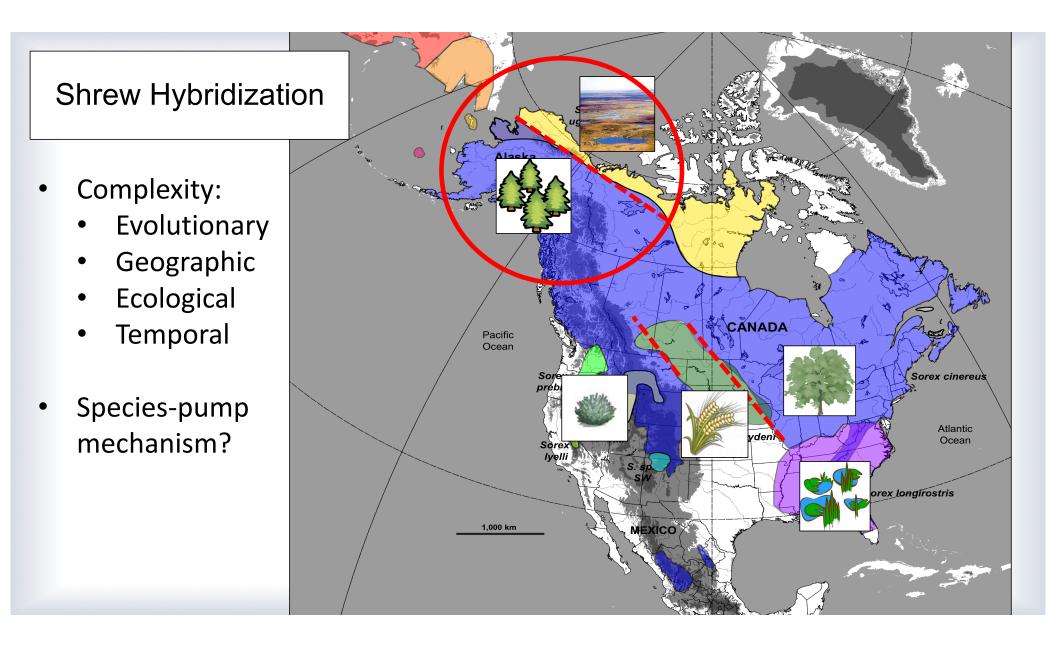




REL (random effects likliehood) Mitogenomes - adaptation

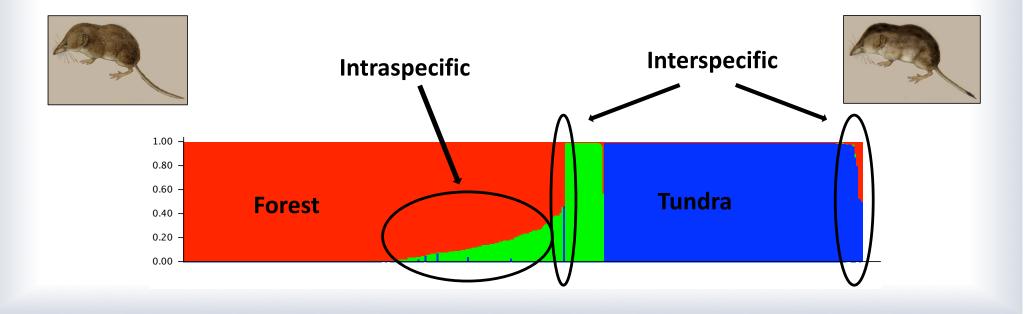






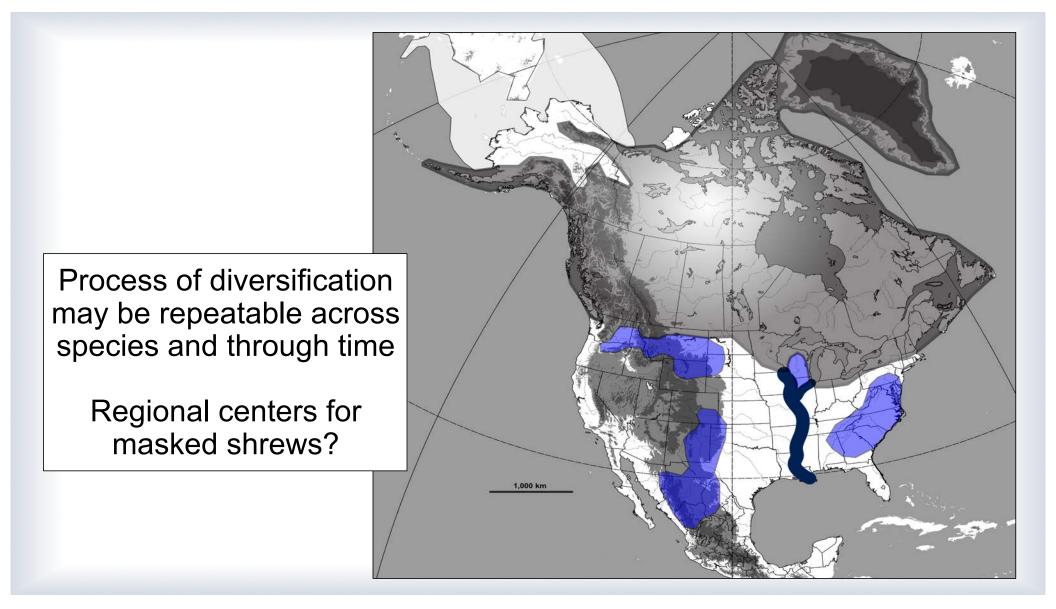
Shrew Hybridization

- Shotgun sequencing (MiSeq) to develop >20 microsatellites
- Rigorous sampling across boreal-tundra contact zone
 - Forest shrew vs. tundra shrew
 - ~400 archived specimens

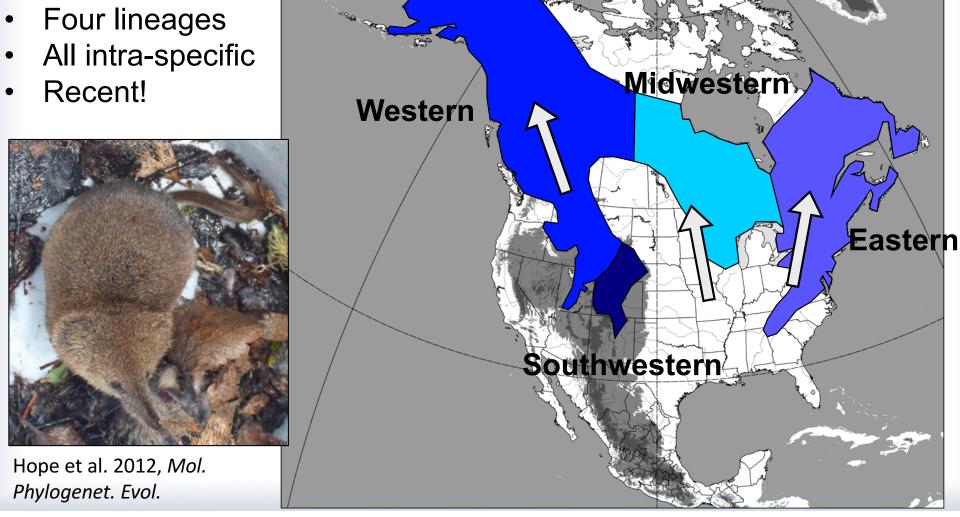


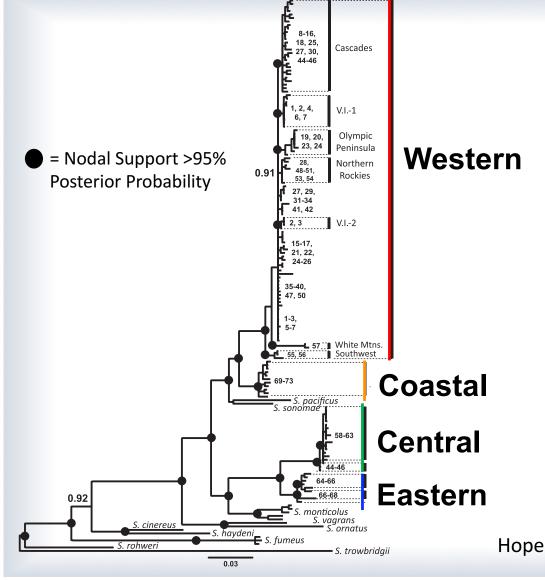
Goals for biodiversity conservation

- 1. Define the taxonomy.
 - Describe species and identify distinct lineages
 - Explore evolutionary processes
- 2. Identify regional centers of diversification
 - Genetic diversity centers/borders over many species?
 - Congruent spatial/temporal history across species?
 - Regional conservation hotspots?
- 3. Resolve connections among associated biodiversity
 - Community ecology
 - Hosts and parasites Co-evolution.
 - Host specificity, host switching



Masked shrews





Other species with same distribution



Water shrew (Sorex palustris)

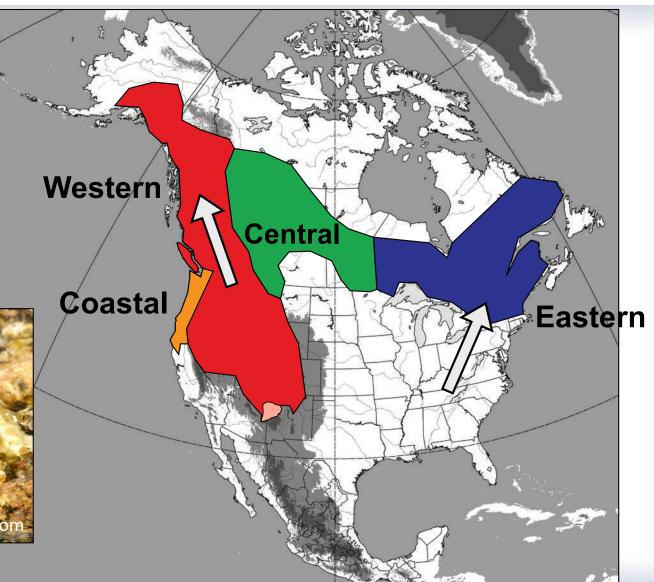
Hope et al. 2014; J. Mamm.

Water shrews

- Four species
- Deep divergence



Hope et al. 2014, J. Mamm.

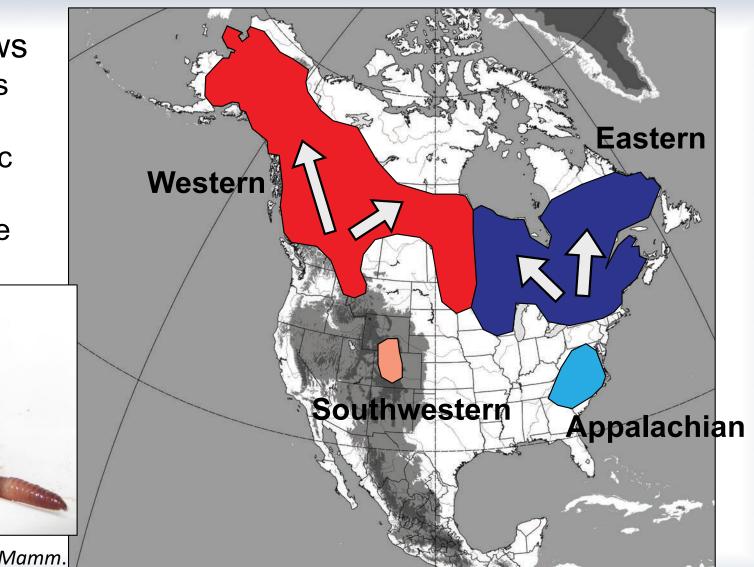


Pygmy shrews

- Two species
- Each with intra-specific lineages
- Intermediate



Hope et al. In Prep for J. Mamm.

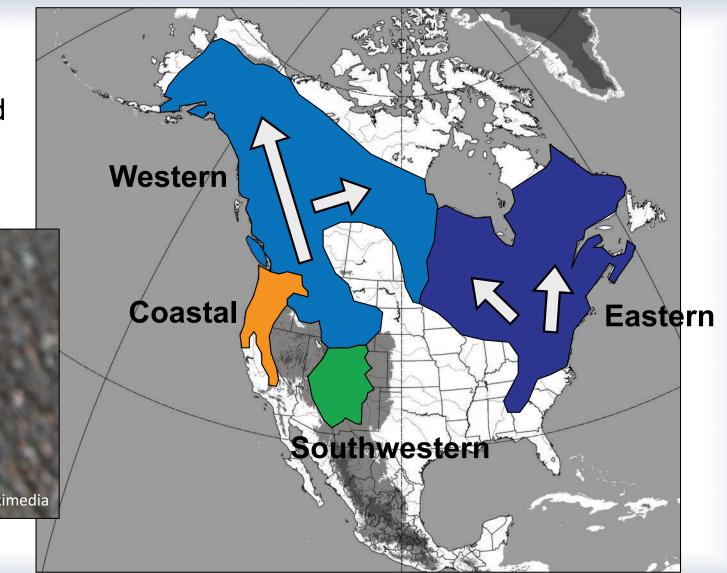


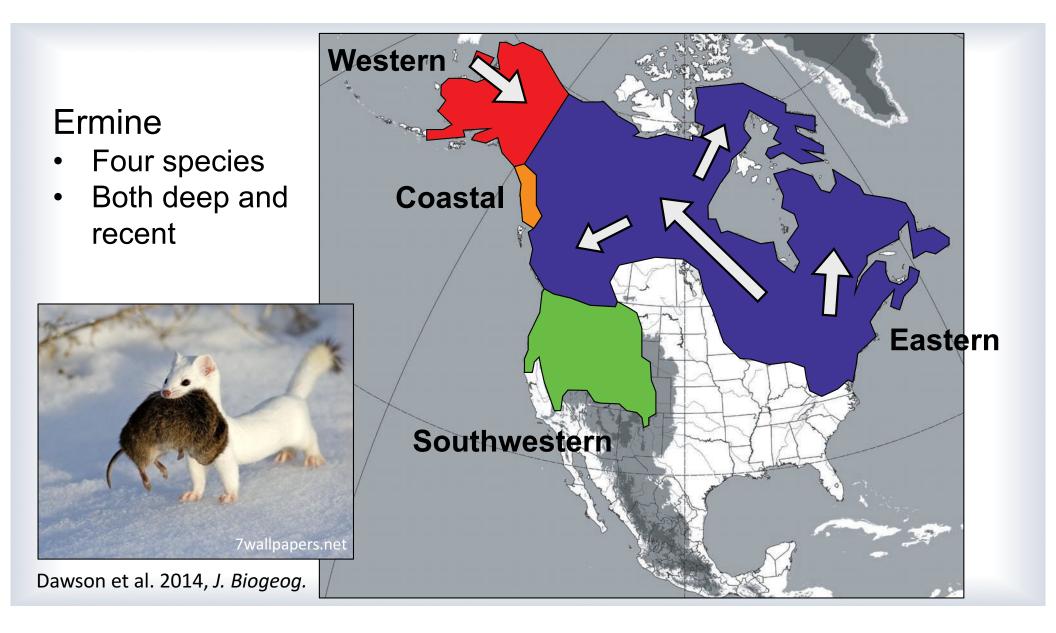
Red squirrels

- Three species
- Both deep and recent divergence



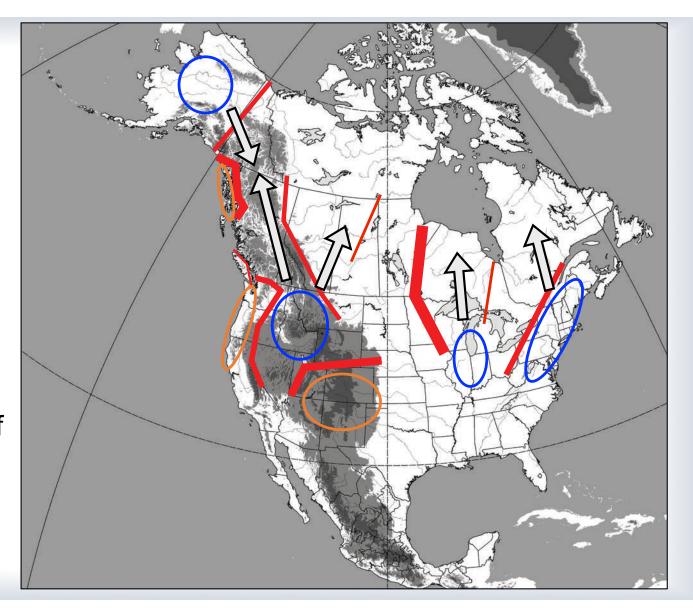
Hope et al. 2016, Mo. Phylogenet. Evol.





Suture Zones

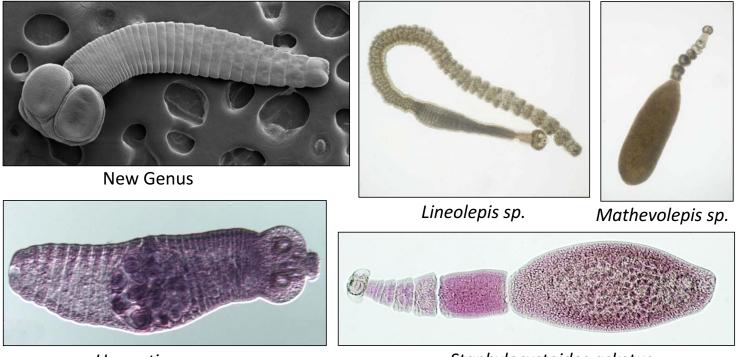
- Multiple areas of contact
- Regionally endemic communities
- Stable regions: high diversity
- Recolonized regions: low diversity
- Pseudocongruence of spatial/temporal diversification.

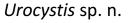


Goals for biodiversity conservation

- 1. Define the taxonomy.
 - Describe species and identify distinct lineages
 - Explore evolutionary processes
- 2. Identify regional centers of diversification
 - Genetic diversity centers/borders over many species?
 - Same spatial/temporal history across species?
 - Regional conservation hotspots?
- 3. Resolve connections among associated biodiversity
 - Community ecology
 - Hosts and parasites Co-evolution.
 - Host specificity, host switching

Interdependent communities



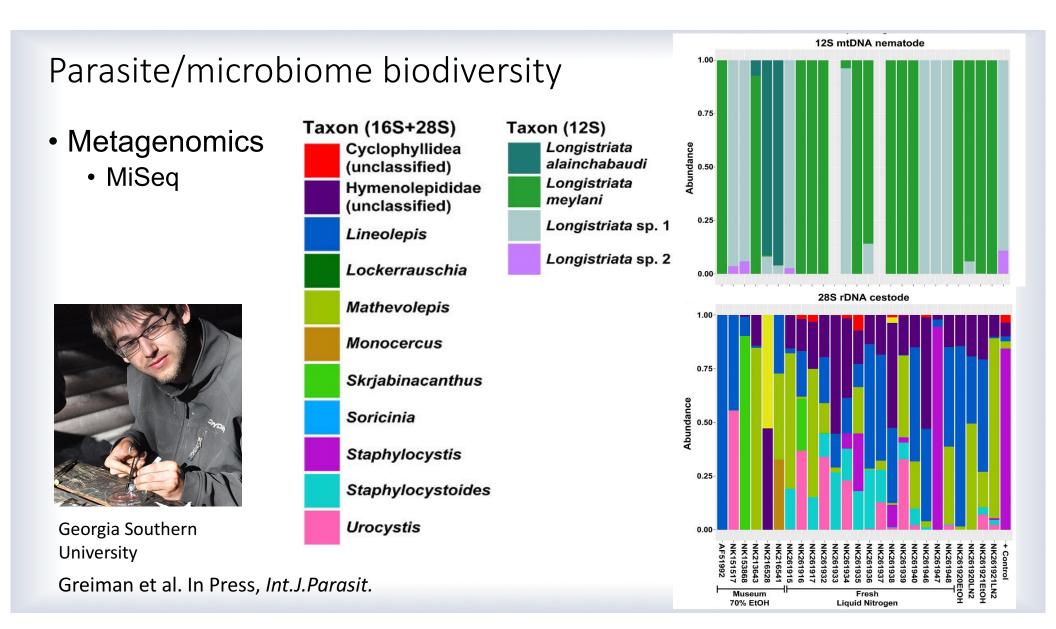


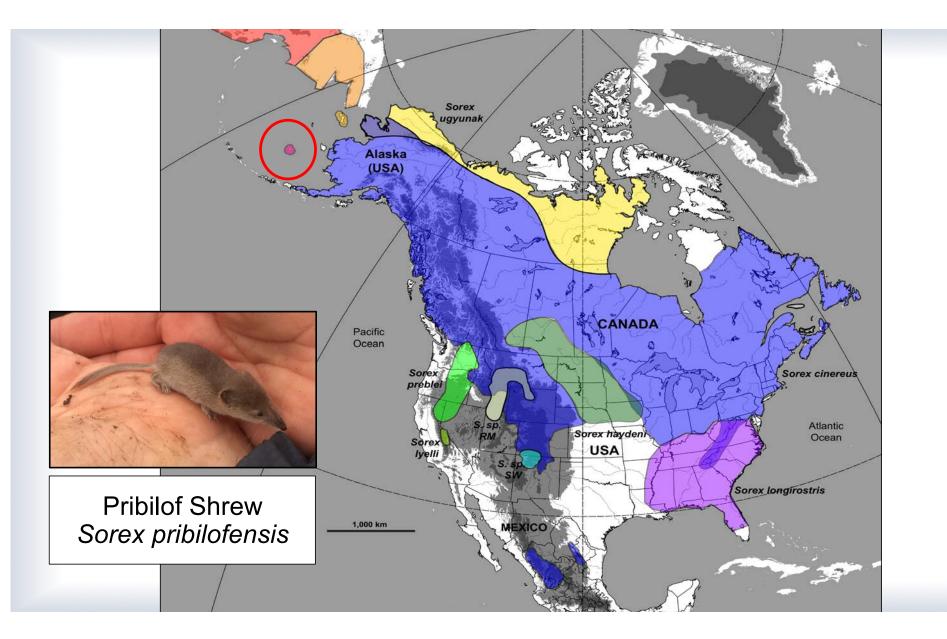
Staphylocystoides asketus

Shrew endoparasites - helminths

- Highly diverse with complex life cycles
- Woefully underexplored!

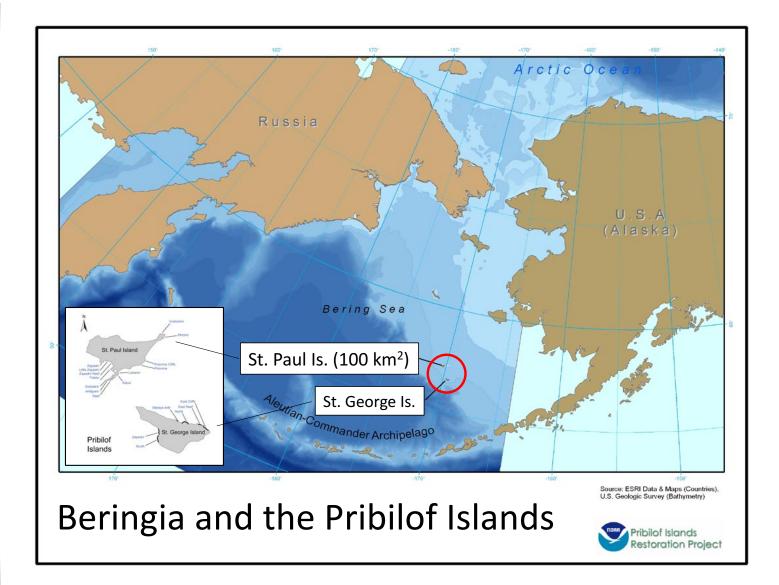
Tkach et al. *in prep (x3)*





St. Paul Island, Alaska.







Predictions for island biodiversity

- 1. Low species richness faunal relaxation
 - Species interdependence \rightarrow cascades (more extreme in north)
 - Low host richness \rightarrow low parasite richness
- 2. Low genetic diversity
 - Small populations \rightarrow periodic bottlenecks
 - Loss of diversity through rapid fixation of alleles (purging)
 - Lowered fitness?
- 3. High endemism
 - Divergence following fragmentation (high drift)
- 4. Host parasite evolutionary concordance?

Mammals of St. Paul Island



Mammals of St. Paul Island



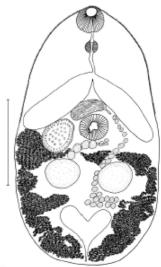
Predictions for island biodiversity

- 1. Low species richness faunal relaxation
 - Species interdependence \rightarrow cascades
 - Low host richness \rightarrow low parasite richness
- 2. Low genetic diversity
 - Small populations \rightarrow periodic bottlenecks
 - Loss of diversity through rapid fixation of alleles (purging)
 - Lowered fitness
- 3. High endemism
 - Divergence following fragmentation (high drift)
- 4. Host parasite evolutionary concordance?

Results – parasite diversity

- Very low Richness 2 species of endoparasite
 - Cestode Lineolepis pribilofensis
 - Trematode *Maritrema* sp. → **Normally in birds!!**
- Very high parasite loads
 - Multiple hundreds of worms per shrew





Karpenko and Dokuchaev 2012

Predictions for island biodiversity

- 1. Low species richness faunal relaxation
 - Species interdependence → cascades
 - Low host richness \rightarrow low parasite richness

2. Low genetic diversity

- Small populations → periodic bottlenecks
- Loss of diversity through rapid fixation of alleles (purging)
- Lowered fitness

3. High endemism

- Divergence following fragmentation (high drift)
- 4. Host parasite evolutionary concordance?

Results – shrew genetic diversity

- Cytochrome b gene (1140bp) for 22 shrews
- Virtually no genetic variability on St. Paul.

Species	Pi	Hd
Sorex pribilofensis	0.0001	0.22
Sorex cinereus (AK)	0.0039	0.95
Sorex cinereus (Southwest)	0.0059	0.89
Sorex portenkoi (Siberia)	0.0005	0.51
Sorex jacksoni (St. Lawrence Is.)	0.0016	0.52
Sorex haydeni (Mid-west)	0.0031	0.86
Sorex preblei (West)	0.0014	0.85

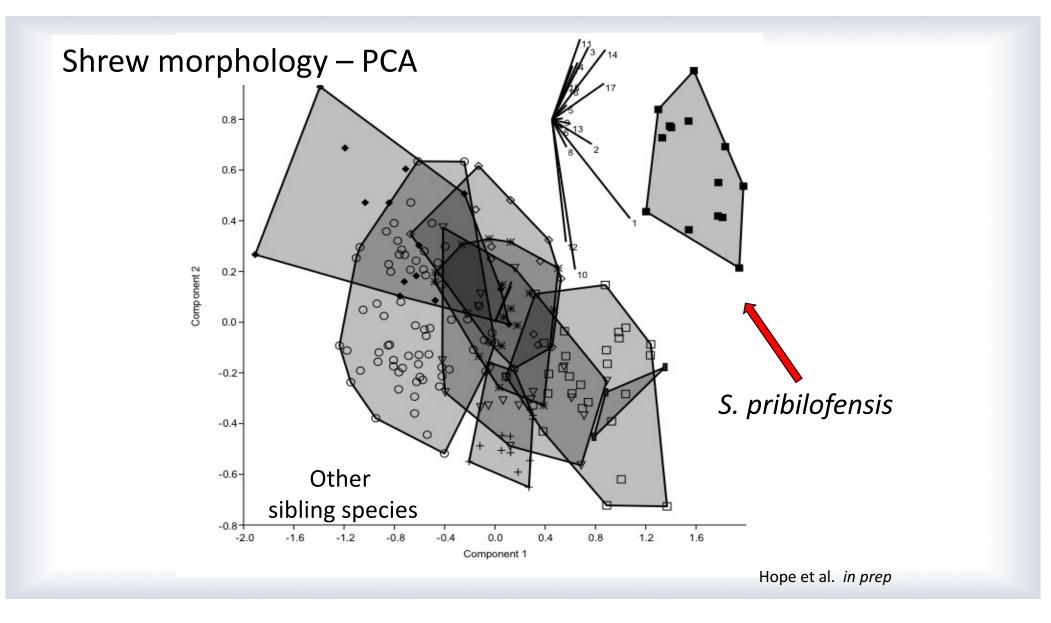
Predictions for island biodiversity

- 1. Low species richness faunal relaxation
 - Species interdependence \rightarrow cascades
 - Low host richness \rightarrow low parasite richness
- 2. Low genetic diversity
 - Small populations \rightarrow periodic bottlenecks
 - Loss of diversity through rapid fixation of alleles

3. High endemism

• Divergence following fragmentation (high drift)

4. Host parasite evolutionary concordance?



Predictions for island biodiversity

- 1. Low species richness faunal relaxation
 - Species interdependence → cascades
 - Low host richness \rightarrow low parasite richness
- 2. Low genetic diversity population demography
 - Small populations \rightarrow periodic bottlenecks
 - Loss of diversity through rapid fixation of alleles
- 3. High endemism
 - Divergence following fragmentation
 - Reflected by phylogeographic histories

4. Host parasite evolutionary concordance?

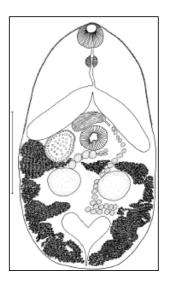
Results – co-evolution?

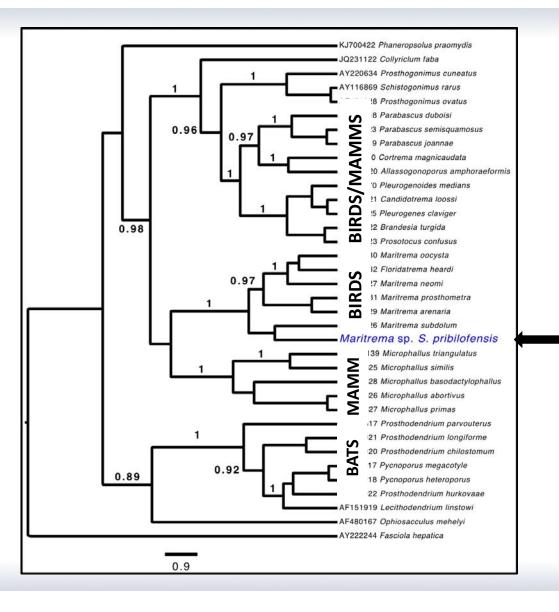
2.0

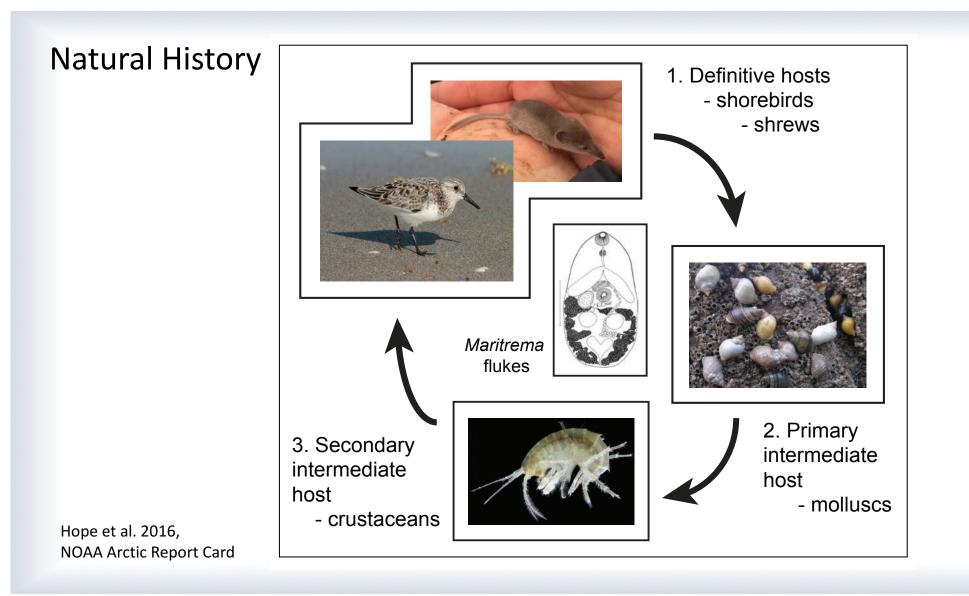
• Shrew cestode with North American origin. Lineolepis S. portenkoi Chukotka Lineolepis Sorex Yukon Lineolepis S. pribilofensis St. Paul 1 Lineolepis S. fumeus West Virginia Nearctic Lineolepis S. cinereus North Dakota 1 Lineolepis S. cinereus North Dakota Lineolepis S. bairdi Oregon Lineolepis S. bairdi Oregon Lineolepis S. cinereus North Dakota Lineolepis S. araneus Czech Repub. 0.81 **Palearctic** 0.93 Lineolepis Sorex Anabar Russ. Lineolepis S. araneus Kazakstan 1 Lineolepis S. araneus Kazakstan 1 Lineolepis Sorex Anabar Russ. 1 Lineolepis Sorex Anabar Russ. Lineolepis S. navigator Yukon Lineolepis Sorex Anabar Russ. 0.92 Lineolepis S. trowbridgii Oregon Neoskrjabinolepis

Results - co-evolution?

 Shrew trematode normally parasitizes birds!







Implications

- 1. Host switching and emerging disease
 - Bird to mammalian
 - Marine to terrestrial
 - Intercontinental movement

2. Low parasite diversity

- High parasite loads
- "Missing biodiversity" dynamics

3. Low shrew genetic diversity

- Loss of adaptive capacity
- Future environmental impacts (raised sea levels!)

Conclusions

- Common species provide windows into ecosystem function
 - Surrogates for rare species
 - Economical
 - Well-represented in museums
 - Often broad geographic focus
 - Resolve regional processes
 - Document connections among species



Thanks To:

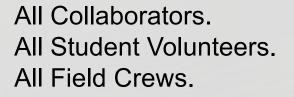
Main Collaborators:

- Joseph A Cook UNM
- Vasyl V Tkach UND
- John R Demboski DMNS
- Sandra L Talbot USGS
- Stephen Greiman GSU





Southwestern Biology



Funding Agencies:

- NSF
- USGS Alaska Science Center
- USFWS
- Aleut Community of St. Paul
- American Society of Mammalogists





Masked shrew "complex"

- 13 closely related species, but...
 - Unresolved genetic relationships
- Very Broad Distributions
 - Selection across environmental gradients?
- Multiple contact zones
 - Hybridization?



Biodiversity conservation...

Conservation of rare species – the "norm"

[CITATION] Survey for the Peñasco least chipmunk (Tamias minimus atristriatus) in the Lincoln National Forest with notes on rodent community assemblages <u>AG Hope</u>, JK Frey - Final report submitted to Lincoln National Forest ..., 2000 ★ 95 Cited by 6 Related articles

2000



Conservation Plan for the Peñasco Least Chipmunk (*Tamias minimus atristriatus*)

Submitted To:

U.S. Fish & Wildlife Service New Mexico Ecological Services Field Office 2105 Osuna Rd. NE Albuquerque, NM 87113

Submitted By:

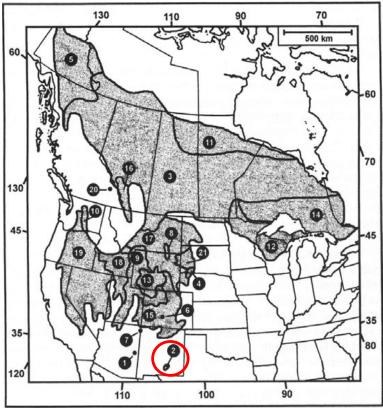
Andrew G Hope, Ph.D. Division of Biology 116 Ackert Hall Kansas State University Manhattan, KS 66506

2017



Facts:

- 1900's very common
- 1930's very common
- 1960's very uncommon
- 1980 virtually gone 2 specimens found
- 2000 virtually gone 2 specimens found
- 2013 virtually gone 2 specimens found
- 2016 petition for listing under the ESA...



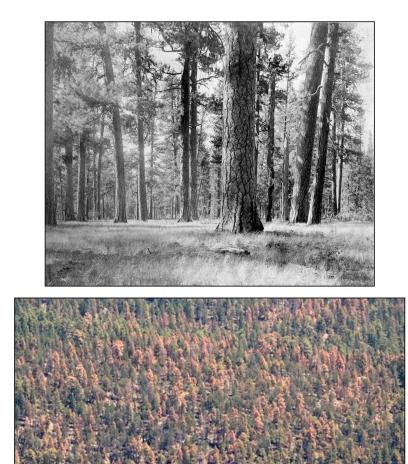
Causes of decline?

- Genetic issues with small populations
- Human land practices grazing, logging, fire suppression
- Drought 1950's
- Disease plague
- Competition
- Hybridization
- All extremely difficult to investigate because this population is virtually gone.

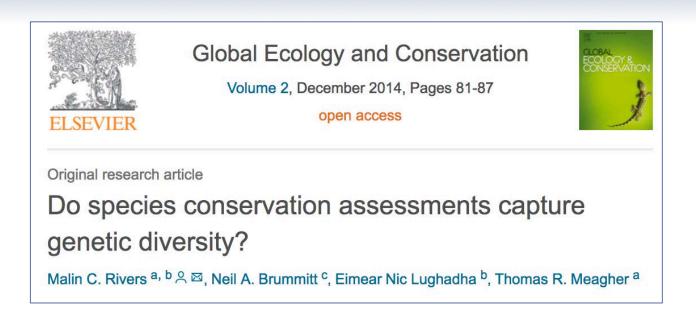


Why should we care about this chipmunk?

- It epitomizes problems:
 - Biodiversity declines can be sudden and fast.
 - Conserving rare taxa is extremely difficult!
 - We normally know very little about rare taxa.
- We should care because:
 - We don't know what role this species fulfilled
 - Indicators of change
 - Need to understand biodiversity connections!!



Biodiversity conservation via evolutionary analysis...

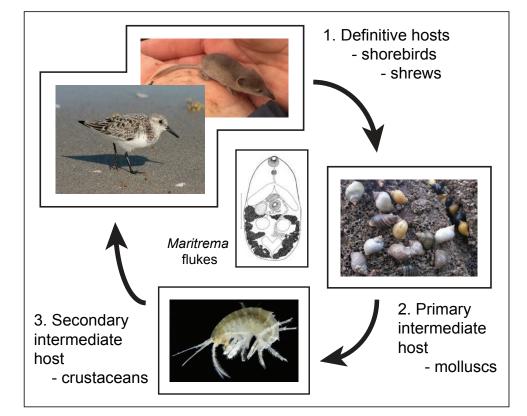


Paraphrasing: "A survey of papers published over the past decade found 8897 papers on the topic "IUCN Red List (population and range size)", and 5505 on "Conservation Genetics"; only 18 (<1%) of these papers included both these topics...

Message: integrating among scientific disciplines is of critical importance for improving biodiversity conservation.

Why is integration among disciplines important?

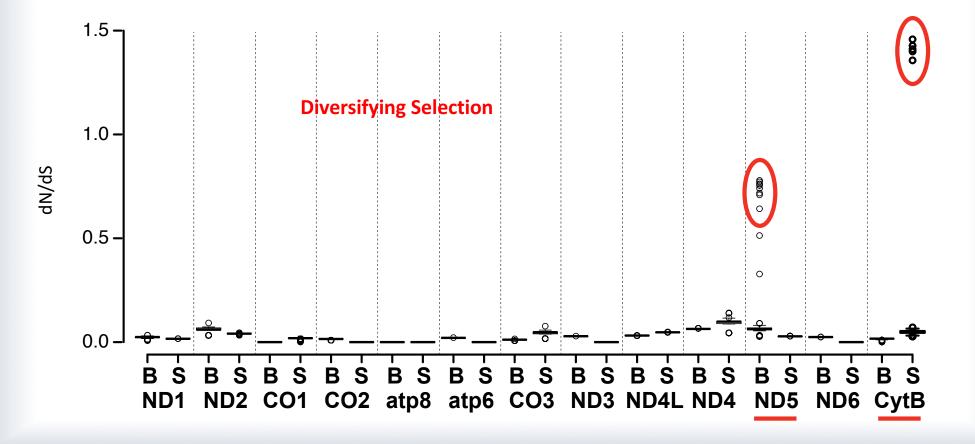
 Maintaining functional ecological systems into the future will benefit from an understanding of both historical and contemporary evolutionary processes that influence fundamental connections among biodiversity.



Biodiversity conservation via evolutionary analysis of *common species*!

Mitogenomes - selection - Zoomed Out...

- Positive Selection in 2 genes



Biggest Threat??

- New biotic associations!
 - New parasite-host relationship
 - New intermediate hosts on island
 - New parasite-environment relationship
 - Ecological fitting
- Terrestrial community reorganization
 - Possible ecological release for snails
 - Vacant niche space and/or lack of competition
- Ecosystem (in)stability
 - Shift in shrew population dynamics?
- Other threats to island?
 - Additional species introductions
 - Rats
 - Other shrews!



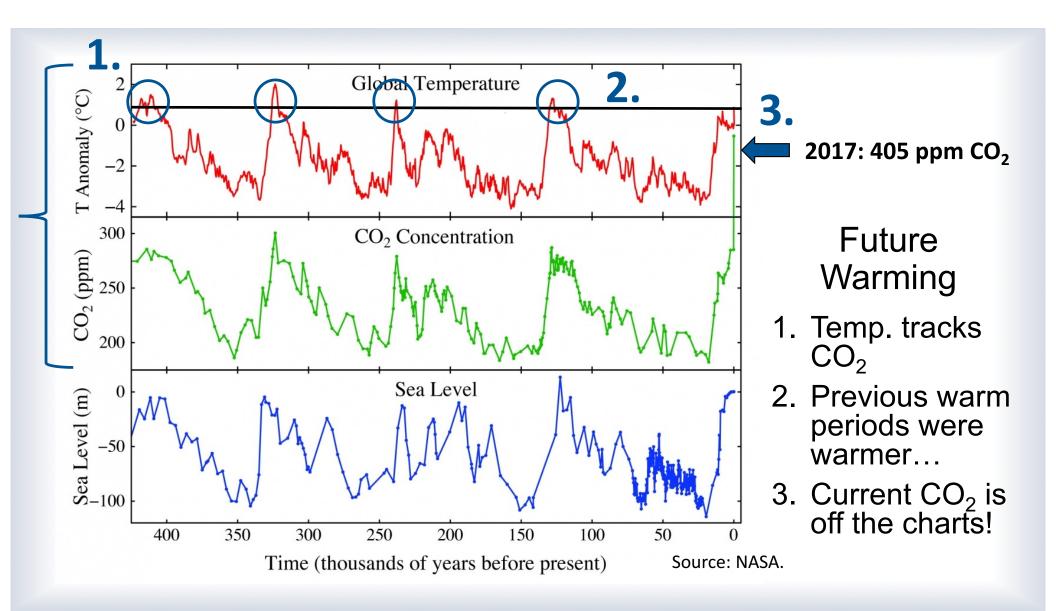






Variation in Intestinal Biodiversity

- Metagenomic methods 12S, 16S, 28S; MiSeq
 - Spatial gradients across host contact zones
 - In-situ temporal change using museum specimens
- Parasite (helminths)
 - Biodiversity discovery
 - Host-parasite co-evolution?
- Microbiomes
 - The next step...

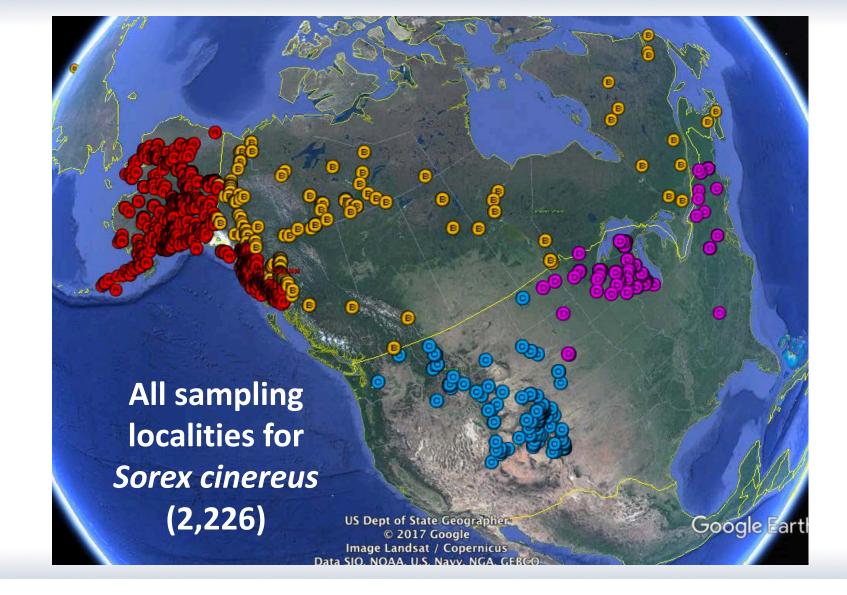


e.g. Sorex cinereus sampling (parentheses indicate average numbers per locality)





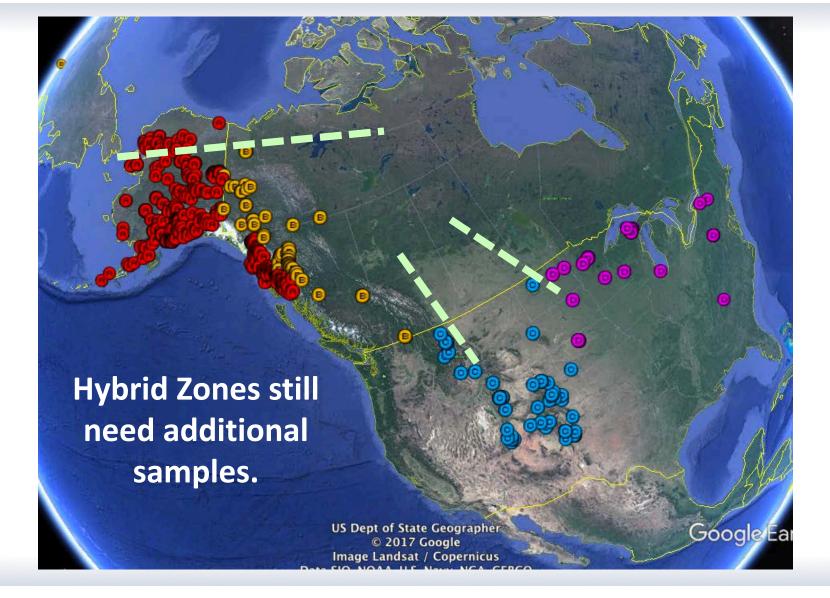
Region	Localities	Total samples	Samples w/ tissues	Samples w/ endoparasites
Alaska	1,391	13,108 (9.4)	11,371 (8.2)	4,687 (3.4)
Canada	291	1,452 (4.9)	905 (3.1)	535 (1.8)
Western US	239	986 (4.1)	445 (1.9)	282 (1.2)
Eastern US	305	1,459 (4.8)	887 (2.9)	172 (0.6)



Sampling localities with intact guts (529; 5 Museums)

> US Dept of State Geographer © 2017 Google Image Landsat / Copernicus

Google/Ear



Summary

- Genomic methods allow for highly integrated ecoevolutionary framework
 - Hybridization, speciation, co-evolution, community assembly
- Combined host-parasite analyses provide greater resolution of generalizable landscape processes
- Understanding interdependency among species...
 - Requires extensive diagnosis of existing biodiversity
 - Requires temporally deep, spatially broad, site intensive, and taxonomically comprehensive sampling

Thanks!

USGS, Alaska Science Center National Science Foundation - Beringian Co-evolution Project Numerous Field Crews!!











THE UNIVERSITY of NEW MEXICO

science for a changing world

KANSAS STATE

UNIVERSITY

NORTH DAKOTA

GEORGIA SOUTHERN

ISGS